

REVIEWS

BIOMARKERS OF HEPATOCELLULAR CARCINOMA: STATUS AND PROSPECTS

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Hepatocellular carcinoma (HCC) also known as hepatocellular cancer is one of the most common and aggressive types of primary malignant liver neoplasms. This type of cancer accounts for up to 90% of all primary liver tumors and is the third leading cause of cancer death worldwide. Despite the advances in modern medicine, diagnostics and treatment of HCC remain challenging, especially in the later stages, when the patient's prognosis significantly worsens and treatment options are very limited. More than half a century has passed since Yu.S. Tatarinov discovered embryo-specific α -globulin in the blood of people with primary liver cancer in 1963, which was later called alpha-fetoprotein (AFP), but unfortunately, the number of specific and sensitive biomarkers for HCC remains very limited. In this regard, many scientific papers are devoted to the search and study of potential HCC biomarkers, which are essential for early diagnostics, prognosis, and development of new therapeutic strategies. Proteomic studies represent one of the promising approaches to investigate both molecular mechanisms of HCC occurrence and HCC biomarkers. Identification of specific protein profiles characteristic of tumor cells can contribute to the identification of new biomarkers that can be used not only for early detection of the disease, but also for monitoring its progression, assessing the response to therapy and predicting the clinical outcome. This review discusses current achievements in the search for potential biomarkers of HCC, as well as the prospects for their clinical use.

Keywords: hepatocellular cancer; biomarkers; protein profiles; proteoforms

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INTRODUCTION

Oncological diseases still remain one of the significant problems of modern global healthcare. According to the World Health Organization (WHO), liver cancer is the sixth most common type of cancer in the world and the third leading cause of cancer deaths [1]. The liver is one of the vital human organs responsible for many body functions, and tumors that develop in it can quickly spread and affect adjacent tissues. Among all types of liver cancer, HCC is the most frequently diagnosed primary liver cancer. According to Rosstat data, during the period from 2010 to 2022 there is a clear increase in percentage of patients with newly diagnosed liver and bile duct cancer in the Russian Federation [2]. More than 850 thousand new cases are diagnosed worldwide every year, and the number of deaths is approaching 800 thousand per year. Despite significant advances in the development of treatment methods, including targeted drugs and immunotherapy, mortality still exceeds the number of primarily registered cases, due to the fairly rapid and asymptomatic development of HCC, and, as a result, diagnostics at later stages [3].

The main factors contributing to the HCC development include genetic and epigenetic changes that gradually disrupt the normal regulation of liver cell

growth and division, leading to their malignant transformation. These include chronic liver damage such as cirrhosis and/or infection with hepatitis B and C viruses [4–6], alcoholic liver disease [7, 8], metabolic disorders (non-alcoholic fatty liver disease (NAFLD) and hemochromatosis), genetic mutations and epigenetic changes [9–12]. According to statistics, the largest number of registered cases of HCC occurs in countries of the Asia-Pacific region. This is primarily due to the low standard of living, high prevalence of chronic viral hepatitis and significant contamination of food products with aflatoxin B1 [13]. In countries with a higher standard of living, there is a tendency for an increase in the HCC incidence in NAFLD patients [14–17]. As the population increases and risk factors become more prevalent, the number of cases is expected to increase by 66% by 2045 [1].

Alpha-fetoprotein (AFP) assay, traditionally used in clinical practice, is characterized by low sensitivity in the early stages of HCC (within 39–65% depending on the baseline level), and its specificity increases up to 90% only at later stages [18, 19]. Moreover, AFP levels can increase not only in HCC, but also in other conditions (ovarian and testicular tumors, pregnancy, etc.) [20–24]. Reliable and specific markers that could effectively identify HCC in the early stages and distinguish it from other liver diseases are needed [25–27]. Since tumor heterogeneity



can affect the effectiveness of individual biomarkers, HCC progression, depending on the etiological factors, can manifest itself in the form of various molecular profiles, thus complicating the development of universal biomarkers [28, 29].

One of the promising approaches for early diagnosis and effective treatment of HCC is the use of proteomics to search for biomarkers that can serve as indicators of the presence and progression of HCC. Unlike the genome, which is relatively static, the proteome, i.e. the set of all proteins in the body, is constantly changing in response to various exogenous and endogenous stimuli [30]. Proteomics is a promising tool for studying cancer, where changes at the protein level (for example, changes in the concentration or modification of proteins) can serve as key indicators of pathological processes.

1. HCC BIOMARKERS USED IN CLINICAL PRACTICE

Modern diagnostics of HCC includes several types of studies: general blood test, serological and instrumental studies (ultrasound (US), computed tomography (CT), and magnetic resonance imaging (MRI)). The main problem in HCC diagnostics is the lack of specific symptoms in the early stages and, as a result, late referral of patients; which has a significant impact on possible limitations of treatment methods. In Russian clinical practice, AFP, the only tumor-specific biomarker is used in combination with non-specific inflammation markers (for example, C-reactive protein (CRP)) (Fig. 1).

In foreign clinical practice, more specific markers are used in addition to AFP; these include the glycosylated form of AFP (AFP-L3) and des-gamma-carboxyprothrombin (DCP), which

are approved biomarkers for diagnostics and monitoring the HCC progression [31]. The use of this combination of biomarkers allows achieving a sensitivity of 94% and a specificity of more than 97% (Fig. 2) [32].

1.1. Alpha-fetoprotein

Alpha-fetoprotein is a glycoprotein that is normally produced in the liver and gall sac of the fetus during intrauterine development. Normally, blood AFP concentrations in adults are low, but they increase in various pathological conditions. The relationship between AFP levels and the HCC development of was first described in 1963 by Tatarinov [33]. AFP is still actively used for screening and diagnostics of HCC, especially in patients with viral hepatitis and cirrhosis of the liver [34]. It is also used to assess evaluate remission/relapse after treatment, but its sensitivity varies between 40–68%, and the concentration may not increase in the early stages or in the presence of small tumors [35]. According to clinical guidelines, serum is used for analysis, and the AFP test result is considered as positive if the AFP level is >100 ng/ml or if it has increased by 7 ng/ml per month based on three consecutive measurements [36].

1.2. Alpha-fetoprotein-L3

AFP-L3 is one of three glycoforms of AFP; it represents the main fraction that binds to *Lens culinaris* agglutinin. Unlike total AFP, which can increase in other liver diseases such as hepatitis or cirrhosis, AFP-L3 is associated primarily with malignant tumors [37]. It demonstrates higher specificity (92–99%) and, according to various studies, the AFP-L3 levels can increase several months before the tumor becomes visible using imaging methods [37, 38].

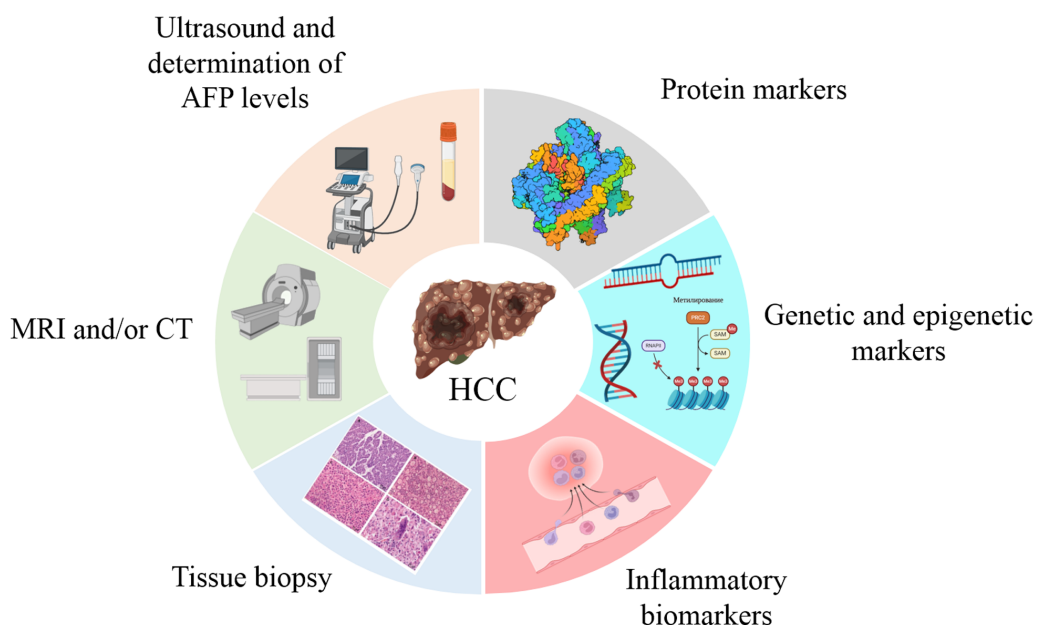


Figure 1. Graphical representation of methods used in HCC diagnostics.

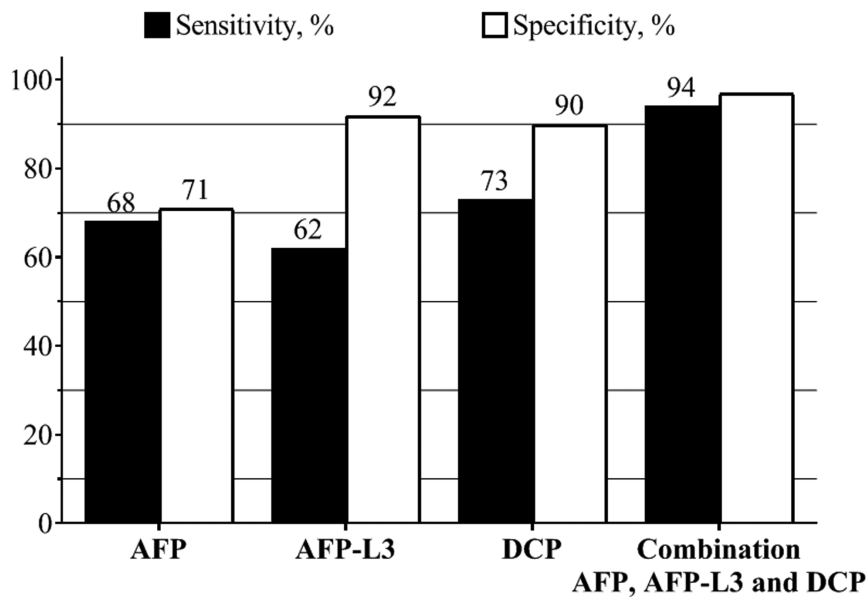


Figure 2. Graphical representation of sensitivity and specificity of HCC biomarkers used in clinical practice.

1.3. Des-gamma-carboxyprothrombin

Des-gamma-carboxyprothrombin is an abnormal form of prothrombin that occurs due to a deficiency of vitamin K-dependent carboxylation. It demonstrates high specificity for HCC, especially in comparison with benign liver diseases, and is used to monitor the response to treatment and assess the risk of recurrence after surgery or other methods of treatment [39, 40].

The levels of the above markers are determined in the blood serum of patients. According to clinical guidelines there are the following reference values: AFP less than 4.7 ng/ml, AFP-L3 less than 10% of the total AFP, DCP less than 7.5 ng/ml.

2. POTENTIAL HCC BIOMARKERS

In recent years, significant progress in molecular research has led to the identification of new potential HCC markers that can significantly improve the diagnostics and monitoring of the disease. In order to be effective in the diagnostics, prognosis and monitoring of treatment of diseases, including HCC, a marker must meet a number of strict criteria, such as high sensitivity and specificity, reproducibility, non-invasiveness, and affordability. Most of the known potential biomarkers have limitations in the context of the listed criteria, so for many decades the search and validation of potential markers has remained a relevant area of research.

If we take into consideration the etiological factors of HCC development, the potential markers can be conditionally subdivided into three main groups: protein biomarkers, genetic and epigenetic biomarkers, and inflammatory biomarkers (Table 1). Many clinical studies have been conducted for many of the listed biomarkers, thus demonstrating both their diagnostic

value and the possibility of using them as a therapeutic target. For example, in April 2024, BioCity (China) began research on a new drug (antibody-drug conjugate) targeting glypican-3 (GPC3). This protein regulates cell growth and differentiation by participating in signaling through interactions with various growth factors (Wnt/ β -catenin signaling pathway) and other molecules, and is also overexpressed in HCC [41]. The first stage of clinical trials is currently underway, and it is expected that GPC3 can be used for a new safe and effective method of HCC treatment [42].

One of the important tasks of modern research in the field of potential markers is the possibility of a minimally invasive method of analysis (blood serum or plasma) known as liquid biopsy. For this type of analysis, the most commonly used methods are, for example, enzyme-linked immunosorbent assay (ELISA), polymerase chain reaction (PCR), next-generation sequencing (NGS), etc. There are a number of disadvantages associated with their use, such as the lack of specific antibodies, the likelihood of false-positive results, and the high cost of research. All this leads to the need of development of new promising approaches for biomarker creation.

Among the potential markers listed in Table 1, miRNAs represent one of the promising group of biomarkers. First of all, this is due to the fact that certain miRNAs can act as oncogenes or tumor suppressors, and, therefore, they be considered as possible therapeutic targets [87–89].

2.1. miR-21

miR-21 is an oncogenic miRNA that suppresses apoptosis and stimulates cell proliferation. Increased expression is observed in HCC and, according to various studies, expression levels correlate with the stage of the disease and metastasis [90–92].

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Table 1. Some potential HCC biomarkers

Biomarker	Description	Type of biomaterial and diagnostic test	Clinical importance	References
<i>Protein biomarkers</i>				
Glypican-3 (GPC3)	Proteoglycan, frequently expressed in HCC cells but absent in normal liver	Blood serum / ELISA	It is used in HCC diagnostics when AFP levels are not elevated. Considered as a possible target for immunotherapy	[38, 43]
Hepatocyte growth factor (HGF)	HGF is associated with HCC progression	Blood serum / ELISA	After liver resection, elevated HGF levels are associated with poor survival prognosis	[44, 45]
Growth differentiation factor-15 (GDF-15)	A cytokine belonging to the family of transforming growth factors	Blood serum / ELISA	Can be used to assess disease severity	[46–48]
Golgi glycoprotein 73 (GP73)	Golgi membrane protein	Blood serum / ELISA	Additional diagnostic marker, more sensitive than AFP	[49, 50]
Vascular endothelial growth factor (VEGF)	Angiogenesis promoting protein	Blood serum / ELISA	Associated with aggressive disease course and poor prognosis. Can be used to assess the effectiveness of therapy	[51, 52]
Proliferation marker protein Ki-67 (Ki-67)	Protein marker of cell proliferation	Tumor biopsates / Immunohistochemical (IHC) or histologic examination	Assesses the rate of tumor growth and its aggressiveness	[53]
Annexin A2 (ANXA2)	A cell membrane associated protein involved in cell adhesion, proliferation and angiogenesis processes	Blood serum / ELISA	It is associated with more aggressive forms of the disease. Considered as a promising biomarker for prognosis and a target for targeted therapy	[54–59]
<i>Genetic and epigenetic markers</i>				
MicroRNA (miRNA)	Small RNA molecules regulating gene expressions: miR-21, miR-26, miR-122, miR-221/miR-222	Blood serum / PCR or NGS	Used in the diagnostics, prognosis, and evaluation of response to treatment	[60–65]
Circulating tumor DNA (ctDNA)	DNA fragments released from tumor cells into the blood circulation	Blood serum / PCR or NGS	Used for early diagnostics and monitoring of HCC recurrence	[66–71]
Beta-catenin (<i>CTNNB1</i>)	Mutations in this gene may be associated with tumor aggressiveness	Tumor tissue / PCR	It is used to identify molecular features and aggressiveness of HCC development	[72–75]
Cellular tumor antigen p53 (<i>TP53</i>)	The gene encoding the tumor suppressor protein p53	Tumor tissue / NGS	Associated with aggressive cancers and unfavorable outcomes	[76–78]
DNA methylation	Epigenetic changes in promoter regions of genes	Plasma, tumor tissue / Bisulfite PCR	Can be used for diagnostics and monitoring of HCC	[79]
<i>Inflammation biomarkers</i>				
C-reactive protein (CRP)	Inflammation biomarker	Blood plasma or serum / Immunochemiluminiscent assay (ICLA)	Increased CRP levels are associated with inflammation and HCC progression	[80–84]
Interleukin-6 (IL-6)	Inflammatory cytokine	Blood serum / ELISA	Increased in HCC and associated with systemic inflammation and poor prognosis	[82, 84–86]

2.2. miR-122

Expression of miR-122 is specific to liver tissue. It acts as a tumor suppressor, playing an important role in the regulation of lipid and glucose metabolism in the liver [93]. A decrease in expression levels correlates with the development of HCC [94], and a number of studies have shown that it can be used as an independent prognostic factor for survival in HCC [95, 96].

2.3. miR-221/miR-222

Highly homologous miRNAs, depending on their expression levels, can act as oncogenes or tumor suppressors. In HCC, miR-221/miR-222 expression levels are significantly elevated; moreover, they correlate with a worse prognosis and increased tumor aggressiveness [97].

2.4. miR-26

miR-26 acts as a tumor suppressor in HCC. Expression levels are significantly reduced and correlate with an increase in the degree of tumor malignancy and a worse survival prognosis [98, 99].

Although the above considered markers have a significant potential in HCC diagnostics, there are a number of limitations associated with their possible use, such as specificity, sensitivity, stability, and clinical significance.

3. PROSPECTS

3.1. Diagnostic Models and Algorithms for HCC Diagnostics

Since new HCC biomarkers have been introduced into clinical practice for a long time, one of the promising approaches is the development of diagnostic algorithms that use existing data on the levels of known clinical markers. In 2014 for HCC diagnostics, Johnson et al. [100] proposed the GALAD (Gender, Age, AFP-L3, AFP, and Des-carboxy-prothrombin) model based on a combination of AFP, AFP-L3, and DCP, including data on gender and age. The study involved 670 patients: 331 HCC patients and 339 patients with chronic liver diseases without signs of oncology as a control. In this groups of patients the model after optimization of sensitivity and specificity showed a prediction accuracy (Area Under Curve, AUC) of 0.88 (95% confidence interval (CI), 0.85–0.91) regardless of the stage of the disease [100].

In 2016, in order to improve the accuracy of HCC diagnostics, the Doylestown Algorithm (DA) using logistic regression was proposed. It includes logarithmically transformed AFP values, as well as age, gender, alkaline phosphatase (ALP), and alanine aminotransferase (ALT) levels [101]. The study was conducted on the basis of electronic data

of 360 patients with liver cirrhosis or HCC that developed in cirrhotic patients. The results have shown that DA increases the overall diagnostic efficiency by 10% as compared to the use of AFP alone. In independent testing performed in three different centers using 2700 patients, the algorithm improved the HCC detection rate from 4% to 20% as compared to AFP alone [101]. This model was later shown to have serious limitations associated with AFP levels below 20.0 ng/ml, and was therefore refined and named as Doylestown Plus. It includes three more potential HCC markers, namely fucosylated alpha-1-antitrypsin (A1AT), fucosylated kininogen (KNG1), and Golgi glycoprotein 73 (GP73). It was shown that DA with the addition of only one marker, KNG1, yielded better results (AUC 0.83) in the overall cohort, but inclusion of GP73 increased the overall individual performance (AUC 0.87) [102].

Based on the GALAD model, Yang et al. [103] proposed an improved version of this model known as GALADUS. It includes not only the GALAD model but also results of ultrasound examination [103]. The use of this model made it possible to achieve an AUC of 0.98 (95% CI, 0.96–0.99), and also to increase the sensitivity and specificity of HCC detection to 95% and 91%, respectively [103].

In 2022, for subsequent assessment of the accuracy of the above algorithms, the data obtained were analyzed in a prospective cohort of patients with Child-Pugh class A or B cirrhosis (408 patients) who were included in the HCC surveillance program between 2004 and 2006. Patient follow-up was terminated if one of the following events occurred: diagnosed HCC, liver transplantation, death, or study discontinuation [104]. Both Doylestown Plus and GALAD algorithms demonstrated higher sensitivity (79% and 72%, respectively) compared to the sensitivity of AFP alone (95% CI, 33.3–90.0). However, it should be noted that this study is limited by small sample sizes and wide CI and requires further investigation [104].

3.2. Development of Test Panels for HCC Diagnostics

Proteomic studies of proteins that determine the proteomic profile for each disease represent a relevant approach to search for potential biomarkers. The human proteome, characterized by heterogeneity and complex composition, consists of polypeptides with or without a certain set of modifications, which are known as protein species or proteoforms [79, 80, 86]. Information on specific fragments of the profile, the so-called protein signatures and the proteoforms that comprise them, can significantly facilitate the search for promising biomarkers. This approach can be used both to search for highly specific proteins secreted by the tumor in small quantities and to develop a panel of several proteins.

To date, there is no standardized panel for HCC screening. The main studies are focused on measuring serum AFP levels and instrumental methods. Therefore, already known HCC markers and their diagnostic improvements are actively studied. The most frequently studied combination of markers includes AFP, AFP-L3, and DCP [105–109]. At the same time, the possibilities of combination with other markers to increase diagnostic sensitivity are also separately considered [50, 110–113].

3.3. Proteoforms as a Source of Biomarkers

In addition to the development of panels of known biomarkers, proteoforms are also actively studied as they can represent highly specific markers of diseases. Proteoforms are formed due to single nucleotide polymorphism (SNP) or alternative splicing or post-translational modifications (PTM). This in turn leads to proteoform diversity and, as a consequence, to diversity of protein functions, their localization, activity, stability and interaction with other molecules. Many PTMs, such as glycosylation, phosphorylation, ubiquitination, etc., are known to be associated with various diseases [114]. A characteristic feature of cancer cells is aberrant glycosylation, which affects different stages of tumor progression. In the case of HCC, the most striking example of a proteoform that has been introduced into clinical practice is the glycosylated form of AFP (AFP-L3) [37].

In the case of other diseases, for example, neurodegenerative ones, the role of beta-amyloid peptide (A β), associated with the rapid progression of Alzheimer's disease (AD), is actively studied. It is known that A β hyperphosphorylation, leading to the formation of amyloid plaques, contributes to the development of AD. A recent study has shown that there are 33 proteoforms of A β associated with rapid progression of AD [115].

In the case of Parkinson's disease (PD), the promising diagnostic and prognostic biomarkers are proteoforms of alpha-synuclein, a protein encoded by the *SNCA* gene. Common PTMs of α -synuclein include phosphorylation of Ser129, eight glycosylated Thr residues (positions 33, 44, 54, 59, 64, 72, 75, 81), and Ser87 [116, 117].

3.4. Bioinformatics Studies

In recent decades, bioinformatics analysis methods have become one of the key links in the search for potential biomarkers. In the context of the rapid growth of data volumes obtained due to modern technologies such as NGS, transcriptomics, proteomics, and metabolomics, bioinformatics research methods represent an effective tool for the systematic analysis of the obtained data. The use of bioinformatics methods allows integrating and analyzing data arrays, identification of new promising markers at various levels of biological organization. Modern databases

(e.g., TCGA or GEO) contain information on genomic, transcriptomic, and proteomic profiles of tumors. Using bioinformatics methods it is possible to analyze these data sets and identify differences in gene/protein expression levels between normal and tumor tissues. This will allow creating a more extensive database of promising HCC markers, which in turn can significantly accelerate the identification of hidden patterns and relationships to create more accurate and personalized approaches to the treatment of malignant tumors [118–120].

CONCLUSIONS

Biomarkers play an important role in the fight against HCC, providing new opportunities for early detection, prognosis and development of targeted therapies. Despite significant progress in the study of HCC biomarkers, many issues remain unresolved. The search for HCC biomarkers is an important direction in improving the diagnostics, prognosis, and disease monitoring. Currently, traditional markers, such as AFP, have their limitations; this emphasizes the need to search for new markers with better diagnostic and prognostic characteristics. Potential biomarkers such as microRNA, changes in DNA methylation, specific protein expression, show significant potential in the early diagnostics of HCC. Such markers can provide a more detailed understanding of the molecular mechanisms of the disease and improve a personalized approach to its treatment. In addition, the introduction of bioinformatics and machine learning methods opens new horizons for data integration and personalization of the treatment. However, despite the promising results of preliminary studies, additional clinical trials are needed to test their reliability and practical value, as well as to develop combined approaches using several markers to improve diagnostic accuracy and prognosis. It is especially important to integrate new biomarkers into existing diagnostic and therapeutic regimens, which may ultimately lead to improved treatment and reduced mortality from this type of cancer.

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COMPLIANCE WITH ETHICAL STANDARDS

This article does not contain any research involving humans or the use of animals as objects.

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

- Bray F, Laversanne M, Sung H, Ferlay J, Siegel R.L., Soerjomataram I, Jemal A. (2024) Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.*, **74**(3), 229–263. DOI: 10.3322/caac.21834
- Aleksandrova G.A., Akhmetzyanova R.R., Golubev N.A., Kirillova G.N., Ogryzko E.V., Oskov Yu.I., Romanenko O.I., Kharkova T.L., Chumarina V.G. (2023) Healthcare in Russia 2023. Federal State Statistics Service (Rosstat), pp. 1–179.
- Park J.-W., Chen M., Colombo M., Roberts L.R., Schwartz M., Chen P.-J., Kudo M., Johnson P., Wagner S., Orsini L.S., Sherman M. (2015) Global patterns of hepatocellular carcinoma management from diagnosis to death: the BRIDGE study. *Liver Int.*, **35**(9), 2155–2166. DOI: 10.1111/liv.12818
- El-Serag H.B. (2012) Epidemiology of viral hepatitis and hepatocellular carcinoma. *Gastroenterology*, **142**(6), 1264–1273e1. DOI: 10.1053/j.gastro.2011.12.061
- Zamor P.J., de Lemos A.S., Russo M.W. (2017) Viral hepatitis and hepatocellular carcinoma: etiology and management. *J. Gastrointest. Oncol.*, **8**(2), 229–242. DOI: 10.21037/jgo.2017.03.14
- Arzumanyan A., Reis H.M.G.P.V., Feitelson M.A. (2013) Pathogenic mechanisms in HBV- and HCV-associated hepatocellular carcinoma. *Nat. Rev. Cancer*, **13**(2), 123–135. DOI: 10.1038/nrc3449
- Ganne-Carrie N., Nahon P. (2019) Hepatocellular carcinoma in the setting of alcohol-related liver disease. *J. Hepatology*, **70**(2), 284–293. DOI: 10.1016/j.jhep.2018.10.008
- Huang D., Mathurin P., Cortez-Pinto H., Loomba R. (2023) Global epidemiology of alcohol-associated cirrhosis and HCC: trends, projections and risk factors. *Nat. Rev. Gastroenterol. Hepatol.*, **20**(1), 37–49. DOI: 10.1038/s41575-022-00688-6
- Schlageter M., Terracciano L., d'Angelo S., Sorrentino P. (2014) Histopathology of hepatocellular carcinoma. *World J. Gastroenterol.*, **20**(43), 15955–15964. DOI: 10.3748/wjg.v20.i43.15955
- Nepomnyashchaya E.M., Shaposhnikov A.V., Yurieva E.A. (2020) Hepatocellular carcinoma: new provisions of the WHO classification, 5th edition, 2019. *Russian Journal of Archive of Pathology*, **82**(6), 36–40. DOI: 10.17116/patol20208206136
- Forner A., Reig M., Bruix J. (2018) Hepatocellular carcinoma. *Lancet*, **391**(10127), 1301–1314. DOI: 10.1016/S0140-6736(18)30010-2
- Tarao K., Nozaki A., Ikeda T., Sato A., Komatsu H., Komatsu T., Taguri M., Tanaka K. (2019) Real impact of liver cirrhosis on the development of hepatocellular carcinoma in various liver diseases-meta-analytic assessment. *Cancer Med.*, **8**(3), 1054–1065. DOI: 10.1002/cam4.1998
- Samant H., Amiri H.S., Zibari G.B. (2020) Addressing the worldwide hepatocellular carcinoma: Epidemiology, prevention and management. *J. Gastrointest. Oncol.*, **12**(2), S361–S373. DOI: 10.21037/jgo.2020.02.08
- Song B.G., Choi S.C., Goh M.J., Kang W., Sinn D.H., Gwak G.-Y., Paik Y.-H., Choi M.S., Lee J.H., Paik S.W. (2023) Metabolic dysfunction-associated fatty liver disease and the risk of hepatocellular carcinoma. *JHEP Reports*, **5**(9), 100810. DOI: 10.1016/j.jhepr.2023.100810
- Talamantes S., Lisjak M., Gilgioni E.H., Llamaza-Torres C.J., Ramos-Molina B., Gurzov E.N. (2023) Non-alcoholic fatty liver disease and diabetes mellitus as growing aetiologies of hepatocellular carcinoma. *JHEP Reports*, **5**(9), 100811. DOI: 10.1016/j.jhepr.2023.100811
- Fang J., Celton-Morizur S., Desdouets C. (2023) NAFLD-related HCC: focus on the latest relevant preclinical models. *Cancers*, **15**(14), 3723. DOI: 10.3390/cancers15143723
- Phoolchand A.G.S., Khakoo S.I. (2024) MASLD and the development of HCC: pathogenesis and therapeutic challenges. *Cancers*, **16**(2), 259. DOI: 10.3390/cancers16020259
- Guse'nov A.Z., Guse'nov T.A. (2016) Modern diagnostics of liver cancer (literature review). *Journal of New Medical Technologies*, **10**(4), 359–377. DOI: 10.12737/23515
- Kushlinsky N.E., Lyubimova N.V. (2016) Tumor markers. General characteristics, clinical significance and recommendations for use. *Poliklinika, Special issue 8*, 62–77.
- Kuo P.-C., Chen S.-C., Shyr Y.-M., Kuo Y.-J., Lee R.-C., Wang S.-E. (2015) Hepatoid carcinoma of the pancreas. *World J. Surg. Oncol.*, **13**(1), 185. DOI: 10.1186/s12957-015-0586-6
- Pedrazzoli P., Rosti G., Soresini E., Ciani S., Secondino S. (2021) Serum tumour markers in germ cell tumours: from to cure. *Crit. Rev. Oncol. Hematol.*, **159**, 103224. DOI: 10.1016/j.critrevonc.2021.103224
- O'Neill A.F., Xia C., Krailo M.D., Shaikh F., Pashankar F.D., Billmire D.F., Olson T.A., Amatruda J.F., Villaluna D., Huang L., Malogolowkin M., Rodriguez-Galindo C., Frazier A.L. (2019) α -Fetoprotein as a predictor of outcome for children with germ cell tumors: a report from the Malignant Germ Cell International Consortium. *Cancer*, **125**(20), 3649–3656. DOI: 10.1002/cncr.32363
- Głowska-Ciemny J., Szmyt K., Kuszarska A., Rzepka R., von Kaisenberg C., Kocylowski R. (2024) Fetal and placental causes of elevated serum alpha-fetoprotein levels in pregnant women. *J. Clin. Med.*, **13**(2), 466. DOI: 10.3390/jcm13020466
- Wang X., Shen C., Yang J., Yang X., Qin S., Zeng H., Wu X., Tang S., Zeng W. (2018) Alpha-fetoprotein as a predictive marker for patients with hepatitis B-related acute-on-chronic liver failure. *Can. J. Gastroenterol. Hepatol.*, **2018**(1), 1232785. DOI: 10.1155/2018/1232785
- Sia D., Villanueva A., Friedman S.L., Llovet J.M. (2017) Liver cancer cell of origin, molecular class, and effects on patient prognosis. *Gastroenterology*, **152**(4), 745–761. DOI: 10.1053/j.gastro.2016.11.048
- Chuang S.-C., la Vecchia C., Boffetta P. (2009) Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer Lett.*, **286**(1), 9–14. DOI: 10.1016/j.canlet.2008.10.040
- Karabork A., Kaygusuz G., Ekin C. (2010) The best immunohistochemical panel for differentiating hepatocellular carcinoma from metastatic adenocarcinoma. *Pathol. Res. Pract.*, **206**(8), 572–577. DOI: 10.1016/j.prp.2010.03.004
- Pinheiro P.S., Jones P.D., Medina H., Cranford H.M., Koru-Sengul T., Bungum T., Wong R., Kobetz E.N., McGlynn K.A. (2024) Incidence of etiology-specific hepatocellular carcinoma: diverging trends and significant heterogeneity by race and ethnicity. *Clin. Gastroenterol. Hepatol.*, **22**(3), 562–571.e8. DOI: 10.1016/j.cgh.2023.08.016

29. Chidambaranathan-Reghupaty S., Fisher P.B., Sarkar D. (2021) Hepatocellular carcinoma (HCC): epidemiology, etiology and molecular classification. *Adv. Cancer Res.*, **149**, 1–61. DOI: 10.1016/bs.acr.2020.10.001
30. Manzoni C., Kia D.A., Vandrovicova J., Hardy J., Wood N.W., Lewis P.A., Ferrari R. (2018) Genome, transcriptome and proteome: the rise of omics data and their integration in biomedical sciences. *Brief. Bioinform.*, **19**(2), 286–302. DOI: 10.1093/bib/bbw114
31. Substantial equivalence determination decision summary assay and instrument combination. Review memorandum K100464. Retrieved May 15, 2024, from: https://www.accessdata.fda.gov/cdrh_docs/reviews/K100464.pdf
32. Behne T., Copur M.S. (2012) Biomarkers for hepatocellular carcinoma. *Int. J. Hepatol.*, **2012**, 859076. DOI: 10.1155/2012/859076
33. Tatarinov Yu.S. (1964) Detection of embryo-specific alpha-globulin in the blood serum of a patient with primary liver cancer. *Voprosy Meditsinskoy Khimii*, **10**(1), 90–91.
34. He C., Peng W., Liu X., Li C., Li X., Wen T.-F. (2019) Post-treatment alpha-fetoprotein response predicts prognosis of patients with hepatocellular carcinoma: a meta-analysis. *Medicine*, **98**(31), e16557. DOI: 10.1097/MD.00000000000016557
35. Hu X., Chen R., Wei Q., Xu X. (2022) The landscape of alpha fetoprotein in hepatocellular carcinoma: where are we? *Int. J. Biol. Sci.*, **18**(2), 536–551. DOI: 10.7150/ijbs.64537
36. Clinical guidelines “Liver Cancer (Hepatocellular)”. Ministry of Health of the Russian Federation. Retrieved September 9, 2023, from: https://oncology-association.ru/wp-content/uploads/2020/09/rak_pecheni.pdf
37. Li D., Mallory T., Satomura S. (2001) AFP-L3: a new generation of tumor marker for hepatocellular carcinoma. *Clin. Chim. Acta*, **313**(1), 15–19. DOI: 10.1016/S0009-8981(01)00644-1
38. Wang J., Wang F., Wang N., Zhang M.-Y., Wang H.-Y., Huang G.-L. (2023) Diagnostic and prognostic value of protein post-translational modifications in hepatocellular carcinoma. *J. Clin. Transl. Hepatol.*, **11**(5), 1192–1200. DOI: 10.14218/JCTH.2022.00006S
39. DCP Test. Retrieved August 05, 2024, from: <https://healthcaresolutions-us.fujifilm.com/products/in-vitro-diagnostics/hcc-risk-biomarkers/dcp-test/>
40. Feng X., Song P., Bie P., Jiang P., Ma K., Li X., Wang S., Wang Z., Tang W., Zheng S. (2016) Des- γ -carboxyprothrombin plasma level in diagnosis of hepatocellular carcinoma in a Chinese population undergoing surgery. *Med. Sci. Monit.*, **22**, 1663–1672. DOI: 10.12659/MSM.895483
41. Devan A.R., Nair B., Pradeep G.K., Alexander R., Vinod B.S., Nath L.R., Calina D., Sharifi-Rad J. (2024) The role of glypican-3 in hepatocellular carcinoma: insights into diagnosis and therapeutic potential. *Eur. J. Med. Res.*, **29**(1), 490. DOI: 10.1186/s40001-024-02073-2
42. BioCity announces FDA clearance of the investigational new drug application for its first-in-class antibody drug conjugate targeting glypican 3 (GPC3). Retrieved June 06, 2024, from: <https://www.biocitypharma.com/news/item?id=43>
43. Shafizadeh N., Kakar S. (2011) Diagnosis of well-differentiated hepatocellular lesions: role of immunohistochemistry and other ancillary techniques. *Adv. Anat. Pathol.*, **18**(6), 438–445. DOI: 10.1097/PAP.0b013e318234abb4
44. Chau G.-Y., Lui W.-Y., Chi C.-W., Chau Y.-P., Li A.-F., Kao H.-L., Wu C.-W. (2008) Significance of serum hepatocyte growth factor levels in patients with hepatocellular carcinoma undergoing hepatic resection. *Eur. J. Surg. Oncol.*, **34**(3), 333–338. DOI: 10.1016/j.ejso.2006.12.007
45. Wang H., Rao B., Lou J., Li J., Liu Z., Li A., Cui G., Ren Z., Yu Z. (2020) The function of the HGF/c-Met axis in hepatocellular carcinoma. *Front. Cell Dev. Biol.*, **8**, 55. DOI: 10.3389/fcell.2020.00055
46. Chen J., Dai W., Zhu C., Liu H., Li Y., Zhang P. (2020) Circulating levels of growth differentiation factor 15 and sex hormones in male patients with HBV-associated hepatocellular carcinoma. *Biomed. Pharmacother.*, **121**, 109574. DOI: 10.1016/j.biopha.2019.109574
47. Li Y., Zhang J., Chen S., Ke Y., Li Y., Chen Y. (2024) Growth differentiation factor 15: emerging role in liver diseases. *Cytokine*, **182**, 156727. DOI: 10.1016/j.cyto.2024.156727
48. Du Y.-N., Zhao J.-W. (2024) GDF15: immunomodulatory role in hepatocellular carcinoma pathogenesis and therapeutic implications. *J. Hepatocell. Carcinoma*, **11**, 1171–1183. DOI: 10.2147/JHC.S471239
49. Zhang X., Wu L.-N., Li X.-Q., Luo X., Liu S.-W., Zhang L., Nawaz S., Ma L.-N., Ding X.-C. (2023) Whether the Golgi protein 73 could be a diagnostic serological marker in hepatocellular carcinoma: a meta analysis. *BMC Gastroenterol.*, **23**(1), 85. DOI: 10.1186/s12876-023-02685-8
50. Wang Y., Wan Y.-J.Y. (2020) Golgi protein 73, hepatocellular carcinoma and other types of cancers. *Liver Res.*, **4**(4), 161–167. DOI: 10.1016/j.livres.2020.09.003
51. Matsui D., Nagai H., Mukozu T., Ogino Y., Sumino Y. (2015) VEGF in patients with advanced hepatocellular carcinoma receiving intra-arterial chemotherapy. *Anticancer Res.*, **35**(4), 2205–2210.
52. Zucman-Rossi J., Villanueva A., Nault J.-C., Llovet J.M. (2015) Genetic landscape and biomarkers of hepatocellular carcinoma. *Gastroenterology*, **149**(5), 1226–1239. DOI: 10.1053/j.gastro.2015.05.061
53. Huang Z., Zhou P., Li S., Li K. (2022) Prediction of the Ki-67 marker index in hepatocellular carcinoma based on dynamic contrast-enhanced ultrasonography with sonazoid. *Insights Imaging*, **13**(1), 199. DOI: 10.1186/s13244-022-01320-6
54. Qiu L.-W., Liu Y.-F., Cao X.-Q., Wang Y., Cui X.-H., Ye X., Huang S.-W., Xie H.-J., Zhang H.-J. (2020) Annexin A2 promotion of hepatocellular carcinoma tumorigenesis via the immune microenvironment. *World J. Gastroenterol.*, **26**(18), 2126–2137. DOI: 10.3748/wjg.v26.i18.2126
55. Zhang H.-J., Yao D.-F., Yao M., Huang H., Wu W., Yan M.-J., Yan X.-D., Chen J. (2012) Expression characteristics and diagnostic value of annexin A2 in hepatocellular carcinoma. *World J. Gastroenterol.*, **18**(41), 5897–5904. DOI: 10.3748/wjg.v18.i41.5897
56. Mohammad H.S., Kurokohchi K., Yoneyama H., Tokuda M., Morishita A., Jian G., Shi L., Murota M., Tani J., Kato K., Miyoshi H., Deguchi A., Himoto T., Usuki H., Wakabayashi H., Izuishi K., Suzuki Y., Iwama H., Deguchi K., Uchida N., Sabat E.A., Arafat U.A., Hassan A.T., El-Sayed A.A., Masaki T. (2008) Annexin A2 expression and phosphorylation are up-regulated in hepatocellular carcinoma. *Int. J. Oncol.*, **33**(6), 1157–1163. DOI: 10.3892/ijo.00000105
57. El-Abd N., Fawzy A., Elbaz T., Hamdy S. (2016) Evaluation of annexin A2 and as potential biomarkers for hepatocellular carcinoma. *Tumor Biol.*, **37**(1), 211–216. DOI: 10.1007/s13277-015-3524-x

58. Zhang H., Yao M., Wu W., Qiu L., Sai W., Yang J., Zheng W., Huang J., Yao D. (2015) Up-regulation of annexin A2 expression predicates advanced clinicopathological features and poor prognosis in hepatocellular carcinoma. *Tumor Biol.*, **36**(12), 9373–9383. DOI: 10.1007/s13277-015-3678-6
59. Tang L., Liu J.-X., Zhang Z.-J., Xu C.-Z., Zhang X.-N., Huang W.-R., Zhou D.-H., Wang R.-R., Chen X.-D., Xiao M.-B., Qu L.-S., Lu C.-H. (2019) High expression of ANXA2 and STAT3 promote progression of hepatocellular carcinoma and predict poor prognosis. *Pathol. Res. Pract.*, **215**(6), 152386. DOI: 10.1016/j.prp.2019.03.015
60. Morishita A., Oura K., Tadokoro T., Fujita K., Tani J., Masaki T. (2021) MicroRNAs in the pathogenesis of hepatocellular carcinoma: a review. *Cancers*, **13**(3), 514. DOI: 10.3390/cancers13030514
61. Gramantieri L., Fornari F., Callegari E., Sabbioni S., Lanza G., Croce C.M., Bolondi L., Negrini M. (2008) MicroRNA involvement in hepatocellular carcinoma. *J. Cell. Mol. Med.*, **12**(6A), 2189–2204. DOI: 10.1111/j.1582-4934.2008.00533.x
62. Xu X., Tao Y., Shan L., Chen R., Jiang H., Qian Z., Cai F., Ma L., Yu Y. (2018) The role of microRNAs in hepatocellular carcinoma. *J. Cancer*, **9**(19), 3557–3569. DOI: 10.7150/jca.26350
63. Singh P., Solanki R., Tasneem A., Suri S., Kaur H., Shah S.R., Dohare R. (2024) Screening of miRNAs as prognostic biomarkers and their associated hub targets across hepatocellular carcinoma using survival-based bioinformatics approach. *J. Genet. Eng. Biotechnol.*, **22**(1), 100337. DOI: 10.1016/j.jgeb.2023.100337
64. Lv Y., Sun X. (2024) Role of miRNA in pathogenesis, diagnosis, and prognosis in hepatocellular carcinoma. *Chem. Biol. Drug Des.*, **103**(1), e14352. DOI: 10.1111/cbdd.14352
65. El Hayek T., Alnaser-Almusa O.A., Alsalameh S.M., Alhalabi M.T., Sabbah A.N., Alshehri E.A., Mir T.A., Mani N.K., Al-Kattan K., Chinnappan R., Yaqinuddin A. (2024) Emerging role of exosomal microRNA in liver cancer in the era of precision medicine; potential and challenges. *Front. Mol. Biosci.*, **11**, 1381789. DOI: 10.3389/fmolb.2024.1381789
66. Bardol T., Pageaux G.-P., Assenat E., Alix-Panabières C. (2024) Circulating tumor DNA clinical applications in hepatocellular carcinoma: current trends and future perspectives. *Clin. Chem.*, **70**(1), 33–48. DOI: 10.1093/clinchem/hvad168
67. Ikeda S., Tsigelny I.F., Skjerveik Å.A., Kono Y., Mendler M., Kuo A., Sicklick J.K., Heestand G., Banks K.C., Talasaz A., Lanman R.B., Lippman S., Kurzrock R. (2018) Next-generation sequencing of circulating tumor DNA reveals frequent alterations in advanced hepatocellular carcinoma. *Oncologist*, **23**(5), 586–593. DOI: 10.1634/theoncologist.2017-0479
68. von Felden J., Craig A.J., Garcia-Lezana T., Labгаа I., Haber P.K., d'Avola D., Asgharpour A., Dieterich D., Bonaccorso A., Torres-Martin M., Sia D., Sung M.W., Tabrizian P., Schwartz M., Llovet J.M., Villanueva A. (2021) Mutations in circulating tumor DNA predict primary resistance to systemic therapies in advanced hepatocellular carcinoma. *Oncogene*, **40**(1), 140–151. DOI: 10.1038/s41388-020-01519-1
69. Li Y., Zheng Y., Wu L., Li J., Ji J., Yu Q., Dai W., Feng J., Wu J., Guo C. (2021) Current status of ctDNA in precision oncology for hepatocellular carcinoma. *J. Exper. Clin. Cancer Res.*, **40**(1), 140. DOI: 10.1186/s13046-021-01940-8
70. Ye Q., Ling S., Zheng S., Xu X. (2019) Liquid biopsy in hepatocellular carcinoma: circulating tumor cells and circulating tumor DNA. *Mol. Cancer*, **18**(1), 114. DOI: 10.1186/s12943-019-1043-x
71. Wu X., Li J., Gassa A., Buchner D., Alakus H., Dong Q., Ren N., Liu M., Odenthal M., Stippel D., Bruns C., Zhao Y., Wahba R. (2020) Circulating tumor DNA as an emerging liquid biopsy biomarker for early diagnosis and therapeutic monitoring in hepatocellular carcinoma. *Int. J. Biol. Sci.*, **16**(9), 1551–1562. DOI: 10.7150/ijbs.44024
72. Rebouissou S., Franconi A., Calderaro J., Letouzé E., Imbeaud S., Pilati C., Nault J.-C., Couchy G., Laurent A., Balabaud C., Bioulac-Sage P., Zucman-Rossi J. (2016) Genotype-phenotype correlation of CTNNB1 mutations reveals different β -catenin activity associated with liver tumor progression. *Hepatology*, **64**(6), 2047–2061. DOI: 10.1002/hep.28638
73. Ding X., Yang Y., Han B., Du C., Xu N., Huang H., Cai T., Zhang A., Han Z.-G., Zhou W., Chen L. (2014) Transcriptomic characterization of hepatocellular carcinoma with CTNNB1 mutation. *PLOS One*, **9**(5), e95307. DOI: 10.1371/journal.pone.0095307
74. Lechrich B.M., Tao J., Liu S., Hirsch T.Z., Yasaka T.M., Cao C., Delgado E.R., Guan X., Lu S., Pan L., Liu Y., Singh S., Poddar M., Bell A., Singhi A.D., Zucman-Rossi J., Wang Y., Monga S.P. (2024) Development of mutated β -catenin gene signature to identify CTNNB1 mutations from whole and spatial transcriptomic data in patients with HCC. *JHEP Reports*, **6**(12), 101186. DOI: 10.1016/j.jhepr.2024.101186
75. Tornesello M.L., Buonaguro L., Tatangelo F., Botti G., Izzo F., Buonaguro F.M. (2013) Mutations in TP53, CTNNB1 and PIK3CA genes in hepatocellular carcinoma associated with hepatitis B and hepatitis C virus infections. *Genomics*, **102**(2), 74–83. DOI: 10.1016/j.ygeno.2013.04.001
76. Yang C., Huang X., Li Y., Chen J., Lv Y., Dai S. (2021) Prognosis and personalized treatment prediction in TP53-mutant hepatocellular carcinoma: an *in silico* strategy towards precision oncology. *Brief. Bioinform.*, **22**(3), bbaa164. DOI: 10.1093/bib/bbaa164
77. Hussain S.P., Schwank J., Staib F., Wang X.W., Harris C.C. (2007) TP53 mutations and hepatocellular carcinoma: insights into the etiology and pathogenesis of liver cancer. *Oncogene*, **26**(15), 2166–2176. DOI: 10.1038/sj.onc.1210279
78. Long J., Wang A., Bai Y., Lin J., Yang X., Wang D., Yang X., Jiang Y., Zhao H. (2019) Development and validation of a TP53-associated immune prognostic model for hepatocellular carcinoma. *eBioMedicine*, **42**, 363–374. DOI: 10.1016/j.ebiom.2019.03.022
79. Fu S., Debes J.D., Boonstra A. (2023) DNA methylation markers in the detection of hepatocellular carcinoma. *Eur. J. Cancer*, **191**, 112960. DOI: 10.1016/j.ejca.2023.112960
80. Ma L.-N., Liu X.-Y., Lu Z.-H., Wu L.-G., Tang Y.-Y., Luo X., Hu Y.-C., Yan T.-T., Wang Q., Ding X.-C., Xie Y. (2017) Assessment of high-sensitivity C-reactive protein tests for the diagnosis of hepatocellular carcinoma in patients with hepatitis B-associated liver cirrhosis. *Oncol. Lett.*, **13**(5), 3457–3464. DOI: 10.3892/ol.2017.5890
81. Sieghart W., Pinter M., Huckle F., Graziadei I., Schöniger-Hekele M., Müller C., Vogel W., Trauner M., Peck-Radosavljevic M. (2013) Single determination of C-reactive protein at the time of diagnosis predicts long-term outcome of patients with hepatocellular carcinoma. *Hepatology*, **57**(6), 2224–2234. DOI: 10.1002/hep.26057

82. Jang J.W., Oh B.S., Kwon J.H., You C.R., Chung K.W., Kay C.S., Jung H.S. (2012) Serum interleukin-6 and C-reactive protein as a prognostic indicator in hepatocellular carcinoma. *Cytokine*, **60**(3), 686–693. DOI: 10.1016/j.cyto.2012.07.017
83. Nagaoka S., Yoshida T., Akiyoshi J., Akiba J., Torimura T., Adachi H., Kurogi J., Tajiri N., Inoue K., Niizeki T., Koga H., Imaizumi T., Kojiro M., Sata M. (2007) Serum C-reactive protein levels predict survival in hepatocellular carcinoma. *Liver Int.*, **27**(8), 1091–1097. DOI: 10.1111/j.1478-3231.2007.01550.x
84. Du J., Huang Z., Zhang E. (2024) Nomograms confirm serum IL-6 and CRP as predictors of immune checkpoint inhibitor efficacy in unresectable hepatocellular carcinoma. *Front. Immunol.*, **15**, 1329634. DOI: 10.3389/fimmu.2024.1329634
85. Tanouti I.-A., Fellah H., Haddaji A., Zerrad C., Tahiri M., Badre W., Nfaoui K., Pineau P., Benjelloun S., Ezzikouri S. (2024) High plasma interleukin-6 level, but not IL-6 gene variants, as a predictive marker for the development of hepatocellular carcinoma in a Moroccan population. *Int. J. Immunogenet.*, **51**(4), 206–216. DOI: 10.1111/iji.12669
86. Porta C., de Amici M., Quaglini S., Paglino C., Tagliani F., Boncimino A., Moratti R., Corazza G.R. (2008) Circulating interleukin-6 as a tumor marker for hepatocellular carcinoma. *Ann. Oncol.*, **19**(2), 353–358. DOI: 10.1093/annonc/mdm448
87. Mallela V.R., Rajtmajerová M., Trailin A., Liška V., Hemminki K., Ambroziewicz F. (2024) miRNA and lncRNA as potential tissue biomarkers in hepatocellular carcinoma. *Non-coding RNA Res.*, **9**(1), 24–32. DOI: 10.1016/j.ncrna.2023.10.010
88. Karakatsanis A., Papaconstantinou I., Gazouli M., Lyberopoulou A., Polymeneas G., Voros D. (2013) Expression of microRNAs, miR-21, miR-31, miR-122, miR-145, miR-146a, miR-200c, miR-221, miR-222, and miR-223 in patients with hepatocellular carcinoma or intrahepatic cholangiocarcinoma and its prognostic significance. *Mol. Carcinog.*, **52**(4), 297–303. DOI: 10.1002/mc.21864
89. Zhang B., Zhu B., Yu J., Liu H., Zhou Y., Sun G., Ma Y., Luan Y., Chen M. (2025) A combined model of six serum microRNAs as diagnostic markers for hepatocellular carcinoma. *Clin. Chim. Acta*, **565**, 119977. DOI: 10.1016/j.cca.2024.119977
90. Qu J., Yang J., Chen M., Cui L., Wang T., Gao W., Tian J., Wei R. (2019) MicroRNA-21 as a diagnostic marker for hepatocellular carcinoma: a systematic review and meta-analysis. *Pak. J. Med. Sci.*, **35**(5), 1466–1471. DOI: 10.12669/pjms.35.5.685
91. Xue X., Li Y., Yao Y., Zhang S., Peng C., Li Y. (2024) A comprehensive review of miR-21 in liver disease: big impact of little things. *Int. Immunopharmacol.*, **134**, 112116. DOI: 10.1016/j.intimp.2024.112116
92. Giordo R., Ahmadi F.A.M., Husaini N.A., Al-Nuaimi N.R.A.M., Ahmad S.M.S., Pintus G., Zayed H. (2024) microRNA 21 and long non-coding RNAs interplays underlie cancer pathophysiology: a narrative review. *Non-coding RNA Res.*, **9**(3), 831–852. DOI: 10.1016/j.ncrna.2024.03.013
93. Chun K.-H. (2022) Molecular targets and signaling pathways of microRNA-122 in hepatocellular carcinoma. *Pharmaceutics*, **14**(7), 1380. DOI: 10.3390/pharmaceutics14071380
94. Tsai W.-C., Hsu P.W.-C., Lai T.-C., Chau G.-Y., Lin C.-W., Chen C.-M., Lin C.-D., Liao Y.-L., Wang J.-L., Chau Y.-P., Hsu M.-T., Hsiao M., Huang H.-D., Tsou A.-P. (2009) MicroRNA-122, a tumor suppressor microRNA that regulates intrahepatic metastasis of hepatocellular carcinoma. *Hepatology*, **49**(5), 1571–1582. DOI: 10.1002/hep.22806
95. Bandiera S., Pfeffer S., Baumert T.F., Zeisel M.B. (2015) miR-122 — a key factor and therapeutic target in liver disease. *J. Hepatology*, **62**(2), 448–457. DOI: 10.1016/j.jhep.2014.10.004
96. Colaianni F., Zelli V., Compagnoni C., Miscione M.S., Rossi M., Vecchiotti D., di Padova M., Alesse E., Zazzeroni F., Tessitore A. (2024) Role of circulating microRNAs in liver disease and HCC: focus on miR-122. *Genes*, **15**(10), 1313. DOI: 10.3390/genes15101313
97. Song Q., An Q., Niu B., Lu X., Zhang N., Cao X. (2019) Role of miR-221/222 in tumor development and the underlying mechanism. *J. Oncology*, **2019**, 7252013. DOI: 10.1155/2019/7252013
98. Li C., Li Y., Lu Y., Niu Z., Zhao H., Peng Y., Li M. (2021) miR-26 family and its target genes in tumorigenesis and development. *Crit. Rev. Oncol. Hematol.*, **157**, 103124. DOI: 10.1016/j.critrevonc.2020.103124
99. Chang L., Li K., Guo T. (2017) miR-26a-5p suppresses tumor metastasis by regulating EMT and is associated with prognosis in HCC. *Clin. Transl. Oncol.*, **19**(6), 695–703. DOI: 10.1007/s12094-016-1582-1
100. Johnson P.J., Pirrie S.J., Cox T.F., Berhane S., Teng M., Palmer D., Morse J., Hull D., Patman G., Kagebayashi C., Hussain S., Graham J., Reeves H., Satomura S. (2014) The detection of hepatocellular carcinoma using a prospectively developed and validated model based on serological biomarkers. *Cancer Epidemiol. Biomarkers Prev.*, **23**(1), 144–153. DOI: 10.1158/1055-9965.EPI-13-0870
101. Wang M., Devarajan K., Singal A.G., Marrero J.A., Dai J., Feng Z., Rinaudo J.A.S., Srivastava S., Evans A., Hann H.-W., Lai Y., Yang H., Block T.M., Mehta A. (2016) The Doylestown algorithm: a test to improve the performance of AFP in the detection of hepatocellular carcinoma. *Cancer Prev. Res.*, **9**(2), 172–179. DOI: 10.1158/1940-6207.CAPR-15-0186
102. Wang M., Sanda M., Comunale M.A., Herrera H., Swindell C., Kono Y., Singal A.G., Marrero J., Block T., Goldman R., Mehta A. (2017) Changes in the glycosylation of kininogen and the development of a kininogen-based algorithm for the early detection of HCC. *Cancer Epidemiol. Biomarkers Prev.*, **26**(5), 795–803. DOI: 10.1158/1055-9965.EPI-16-0974
103. Yang J.D., Addissie B.D., Mara K.C., Harmsen W.S., Dai J., Zhang N., Wongjarupong N., Ali H.M., Ali H.A., Hassan F.A., Lavu S., Cvinar J.L., Giama N.H., Moser C.D., Miyabe K., Allotey L.K., Algeciras-Schimmich A., Theobald J.P., Ward M.M., Nguyen M.H., Befeler A.S., Reddy K.R., Schwartz M., Harnois D.M., Yamada H., Srivastava S., Rinaudo J.A., Gores G.J., Feng Z., Marrero J.A., Roberts L.R. (2019) GALAD score for hepatocellular carcinoma detection in comparison with liver ultrasound and proposal of GALADUS score. *Cancer Epidemiol. Biomarkers Prev.*, **28**(3), 531–538. DOI: 10.1158/1055-9965.EPI-18-0281
104. Singal A.G., Tayob N., Mehta A., Marrero J.A., Jin Q., Lau J., Parikh N.D. (2022) Doylestown Plus and GALAD demonstrate high sensitivity for HCC detection in patients with cirrhosis. *Clin. Gastroenterol. Hepatol.*, **20**(4), 953–955.e2. DOI: 10.1016/j.cgh.2021.04.018
105. Pai S., Parikh N.D. (2024) Novel blood-based biomarkers for HCC. *Curr. Hepatol. Rep.*, **23**(1), 174–184. DOI: 10.1007/s11901-023-00626-3

106. Norman J.S., Li P.J., Kotwani P., Shui A.M., Yao F., Mehta N. (2023) AFP-L3 and DCP strongly predict early hepatocellular carcinoma recurrence after liver transplantation. *J. Hepatology*, **79**(6), 1469–1477. DOI: 10.1016/j.jhep.2023.08.020
107. Oka H., Saito A., Ito K., Kumada T., Satomura S., Kasugai H., Osaki Y., Seki T., Kudo M., Tanaka M. (2001) Multicenter prospective analysis of newly diagnosed hepatocellular carcinoma with respect to the percentage of *Lens culinaris* agglutinin-reactive alpha-fetoprotein. *J. Gastroenterol. Hepatol.*, **16**(12), 1378–1383. DOI: 10.1046/j.1440-1746.2001.02643.x
108. Cheng J., Wang W., Zhang Y., Liu X., Li M., Wu Z., Liu Z., Lv Y., Wang B. (2014) Prognostic role of pre-treatment serum AFP-L3% in hepatocellular carcinoma: systematic review and meta-analysis. *PLOS One*, **9**(1), e87011. DOI: 10.1371/journal.pone.0087011
109. Zinkin N.T., Grall F., Bhaskar K., Otu H.H., Spentzos D., Kalmowitz B., Wells M., Guerrero M., Asara J.M., Libermann T.A., Afdhal N.H. (2008) Serum proteomics and biomarkers in hepatocellular carcinoma and chronic liver disease. *Clin. Cancer Res.*, **14**(2), 470–477. DOI: 10.1158/1078-0432.CCR-07-0586
110. Cummings J., Apostolova L., Rabinovici G.D., Atri A., Aisen P., Greenberg S., Hendrix S., Selkoe D., Weiner M., Petersen R.C., Salloway S. (2023) Lecanemab: appropriate use recommendations. *J. Prev. Alzheimers Dis.*, **10**(3), 362–377. DOI: 10.14283/jpad.2023.30
111. Cheng K., Shi J., Liu Z., Jia Y., Qin Q., Zhang H., Wan S., Niu Z., Lu L., Sun J., Xue J., Lu C., Wei X., Guo L., Zhang F., Zhou D., Tang Y., Hu Y., Huang Y., Chen Y., Lau W.Y., Cheng S., Liu S. (2020) A panel of five plasma proteins for the early diagnosis of hepatitis B virus-related hepatocellular carcinoma in individuals at risk. *eBioMedicine*, **52**, 102638. DOI: 10.1016/j.ebiom.2020.102638
112. Zhao S., Long M., Zhang X., Lei S., Dou W., Hu J., Du X., Liu L. (2020) The diagnostic value of the combination of Golgi protein 73, glypican-3 and alpha-fetoprotein in hepatocellular carcinoma: a diagnostic meta-analysis. *Ann. Transl. Med.*, **8**(8), 536. DOI: 10.21037/atm.2020.02.89
113. Cao W.-Q., Jiang B.-Y., Huang J.-M., Zhang L., Liu M.-Q., Yao J., Wu M.-X., Zhang L.-J., Kong S.-Y., Wang Y., Yang P.-Y. (2019) Straightforward and highly efficient strategy for hepatocellular carcinoma glycoprotein biomarker discovery using a nonglycopeptide-based mass spectrometry pipeline. *Anal. Chem.*, **91**(19), 12435–12443. DOI: 10.1021/acs.analchem.9b03074
114. Xu H., Wang Y., Lin S., Deng W., Peng D., Cui Q., Xue Y. (2018) PTMD: a database of human disease-associated post-translational modifications. *Genomics Proteomics Bioinformatics*, **16**(4), 244–251. DOI: 10.1016/j.gpb.2018.06.004
115. Noor A., Zafar S., Shafiq M., Younas N., Siegert A., Mann F.A., Kruss S., Schmitz M., Dihazi H., Ferrer I., Zerr I. (2022) Molecular profiles of amyloid- β proteoforms in typical and rapidly progressive Alzheimer's disease. *Mol. Neurobiol.*, **59**(1), 17–34. DOI: 10.1007/s12035-021-02566-9
116. Pons M.-L., Loftus N., Vialaret J., Moreau S., Lehmann S., Hirtz C. (2022) Proteomics challenges for the assessment of synuclein proteoforms as clinical biomarkers in Parkinson's disease. *Front. Aging Neurosci.*, **14**, 818606. DOI: 10.3389/fnagi.2022.818606
117. Levine P.M., Galesic A., Balana A.T., Mahul-Mellier A.-L., Navarro M.X., de Leon C.A., Lashuel H.A., Pratt M.R. (2019) α -Synuclein O-GlcNAcylation alters aggregation and toxicity, revealing certain residues as potential inhibitors of Parkinson's disease. *Proc. Natl. Acad. Sci. USA*, **116**(5), 1511–1519. DOI: 10.1073/pnas.1808845116
118. Luo J.-P., Wang J., Huang J.-H. (2021) CDKN2A is a prognostic biomarker and correlated with immune infiltrates in hepatocellular carcinoma. *Biosci. Rep.*, **41**(10), BSR20211103. DOI: 10.1042/BSR20211103
119. Li Y., Chen R., Yang J., Mo S., Quek K., Kok C.H., Cheng X.-D., Tian S., Zhang W., Qin J.-J. (2020) Integrated bioinformatics analysis reveals key candidate genes and pathways associated with clinical outcome in hepatocellular carcinoma. *Front. Genet.*, **11**, 814. DOI: 10.3389/fgene.2020.00814
120. Jiang C.H., Yuan X., Li J.F., Xie Y.F., Zhang A.Z., Wang X.L., Yang L., Liu C.X., Liang W.H., Pang L.J., Zou H., Cui X.B., Shen X.H., Qi Y., Jiang J.F., Gu W.Y., Li F., Hu J.M. (2020) Bioinformatics-based screening of key genes for transformation of liver cirrhosis to hepatocellular carcinoma. *J. Transl. Med.*, **18**(1), 40. DOI: 10.1186/s12967-020-02229-8

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БИОМАРКЕРЫ ГЕПАТОЦЕЛЛЮЛЯРНОГО РАКА: СОСТОЯНИЕ И ПЕРСПЕКТИВЫ

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Гепатоцеллюлярная карцинома или гепатоцеллюлярный рак (ГЦР) представляет собой один из наиболее распространённых и агрессивных видов первичных злокачественных новообразований печени. Этот вид рака составляет до 90% всех первичных опухолей печени и занимает третье место среди причин смертности от онкологических заболеваний в мире. Несмотря на достижения современной медицины, диагностика и лечение ГЦР остаются сложными задачами, особенно на поздних стадиях, когда прогноз для пациента значительно ухудшается, а вариант выбора лечения весьма ограничен. С момента обнаружения Ю.С. Татариновым в 1963 году эмбрионспецифического α -глобулина в крови людей, болеющих первичным раком печени, впоследствии получившего название альфа-фетопротеин (АФП), прошло более половины столетия, но, к сожалению, количество специфичных и чувствительных биомаркеров для ГЦР остаётся весьма ограниченным. В связи с этим, многие научные работы посвящены поиску и изучению потенциальных биомаркеров ГЦР, имеющих существенное значение для ранней диагностики, прогноза и разработки новых терапевтических стратегий. Одним из перспективных подходов к изучению молекулярных механизмов возникновения ГЦР и поиску биомаркеров являются протеомные исследования. Выявление специфических белковых профилей, характерных для опухолевых клеток, может способствовать идентификации новых биомаркеров, которые могут быть использованы не только для раннего выявления заболевания, но и для мониторинга его прогрессирования, оценки ответа на терапию и предсказания клинического исхода. В данном обзоре рассмотрены современные достижения по поиску потенциальных биомаркеров ГЦР, а также перспективы их клинического использования.

Полный текст статьи на русском языке доступен на сайте журнала (<http://pbmc.ibmc.msk.ru>).

Ключевые слова: гепатоцеллюлярный рак; биомаркеры; белковые профили; протеоформы

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